# Protocol of P450-Glo™ CYP2D6 Assay for High-throughput Screening

**DOCUMENT:** P450-Glo<sup>TM</sup> CYP2D6\_TOX21\_SLP\_Version1.0

TITLE: Protocol of P450-Glo<sup>TM</sup> CYP2D6 Assay for High-throughput Screening

#### **ASSAY RFERENCES:**

Assay Target	Cell Lines	Species	Tissue of Origin	Assay Readout	Assay Provider	Toxicity Pathway
CYP2D6	N/A	Human	Enzyme	Luminescence	Promega Corporation	Cell-free cytochrome P450

# **QUALITY CONTROL PRECAUTIONS:**

- 1. Avoid repeated freeze-thaw cycles of substrate and enzyme.
- 2. Dispense the unused Luciferin-ME EGE and CYP2D6 membranes into single-use aliquots and store at -70°C.
- 3. Keep the mixtures on ice during assay run.

#### **MATERIALS and INSTRUMENTS:**

Supplies/Medium/Reagent	Manufacturer	Vender/Catalog Number	
-P450-Glo™ CYP2D6 Screening System	-Promega	-Promega/V9890	
-Quinidine (Positive control compound)	-Sigma Aldrich	-Sigma Aldrich/Q3625	
-1536-well medium binding white solid plates	-Greiner Bio-one	-Greiner Bio-one/789175-F	
-BioRAPTR FRD	-Beckman Coulter	-Beckman Coulter	
-Pintool station	-Wako	-Wako	
-ViewLux plate reader	-Perkin Elmer	-Perkin Elmer	

### **PROCEDURE:**

#### 1. Enzyme-substrate mixture:

- i. Luciferin-Free water = 18.93uL (this addition is to make up the volume)
- ii. Potassium phosphate buffer, 1M = 5.0uL
- iii. 10mM Luciferin-ME EGE = 0.15uL (10mM Luciferin-ME EGE solution was prepared by dissolving 900ug of Luciferin-ME EGE in 265ul of acetonitrile and mix vigorously)
- iv. CYP2D6 membranes = 0.25uL
- v. 0.2% BSA (7.5%BSA stock) = 0.67uL
- vi. Final volume = 25uL

# 2. NADPH regeneration solution:

- i. Luciferin-Free water = 22.0uL
- ii. Solution A = 2.5uL
- iii. Solution B = 0.5uL
- iv. Final Volume = 25.0uL

## 3. Assay Protocol

- i. Two uL of enzyme-substrate mixture was dispensed dispenser in 1536-well medium-binding white solid plates using BioRAPTR dispenser.
- ii. The positive control and test compounds were transferred at 23nL to 1-4 and 5-48 columns of the assay plates respectively using Pintool station.

Positive control plate format: Columns 1: two-fold sixteen-point titration starting at 250uM (final = 1.44uM) duplicates; Columns 2 & 3 (top halves): 125uM (final = 0.72uM) and 62.5uM (0.36uM) Quinidine respectively; Columns 2 & 3 (bottom halves) and 4: DMSO.

Control used is Quinidine [250uM made in DMSO (final = 1.44uM)]

- iii. The assay plates were incubated for 10min at room temperature.
- iv. The reaction was initiated through the addition of 2uL NADPH regeneration solution using BioRAPTR dispenser.
- v. The assay plates were incubated for 60min at room temperature to allow the reaction to continue before it was stopped with a detection reagent.
- vi. Four uL of detection reagent was added using BioRAPTR dispenser.
- vii. The assay plates were incubated for 20min at room temperature.
- viii. The luminescence intensity was quantified using ViewLux plate reader (Exposure time: \_\_\_\_\_\_\_ 10sec).

**ASSAY** 

#### 

**PERFORMANCE:** 

*CV values shown represent average of DMSO column from each plate.