Protocol of AhR HepG2 Cell-based Assay for High-throughput Screening

DOCUMENT: AhR_TOX21_SLP_Version1.0

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ASSAY RFERENCES:

| Assay Target | Cell Lines | Species | Tissue of Origin | Assay Readout | Assay Provider | Toxicity Pathway |
|----------------------------|---------------|---------|--------------------------|---------------------|-------------------|---------------------|
| Human AhR (Recombinant) | HepG2 | Human | Hepatocellular carcinoma | Luciferase reporter | Denison Lab | NR signaling |

QUALITY CONTROL PRECAUTIONS:

MATERIALS and INSTRUMENTS:

| Manufacturer | Vender/Catalog Number | |
|------------------------|--|--|
| -Invitrogen | -12561 | |
| -Atlanta Biologicals | -S11150 | |
| -Invitrogen | -10131-027. | |
| -Invitrogen | -25300 | |
| -Invitrogen | -14190 | |
| -AMRESCO | -KD Medical, RGE-3070 | |
| -Invitrogen | -12648 | |
| -Greiner | - | |
| -Sigma Aldrich | - | |
| -Kalypsis | - | |
| -PerkinElmer | - | |
| -Sorvall legend XTR | -Thermo Fisher Science 75004520 | |
| -sigma | -88000 | |
| -Promega | -E6130 | |
| -Promega | -G6082 | |
| | -Invitrogen -Atlanta Biologicals -Invitrogen -Invitrogen -Invitrogen -AMRESCO -Invitrogen -Greiner -Sigma Aldrich -Kalypsis -PerkinElmer -Sorvall legend XTR -sigma -Promega | |

PROCEDURE:

1. Cell handling:

1.1. Media Required:

| Component | Growth Medium | Assay Medium | Thaw Medium | Freezing Medium |
|--|------------------|-----------------|----------------|--------------------|
| -Alpha MEM | -90% | -90% | -90% | - |
| -Premium FBS | -10% | -10% | -10% | - |
| -G418 | -400mg/mL | - | - | - |
| -Recovery cell culture freezing medium | - | - | - | -100% |

1.2. Thawing method

- 1.2.1 -place 14mL of pre-warmed thaw media into T75 flask
- 1.2.2 -remove vial of cells to be thawed from -140 and thaw rapidly by placing in water bath with gentle agitation for 1-2min
- 1.2.3 -Wipe vial with 70% ethanol before opening in biological safety cabinet
- 1.2.4 -transfer vial contents dropwise into 10mL of thaw medium in 15mL conical tube
- 1.2.5 -centrifuge cells at 1000 rpm for 4min
- 1.2.6 -Resuspend and transfer contents into T75 flask containing thaw medium and transfer flask into incubator
- 1.2.7 -switch to growth media at first passage

1.3. Propagation method

- 1.3.1 -aspirate media, rinse once with dPBS, add 0.25% trypsin/EDTA and swirl to coat flask evenly
- 1.3.2 -add equal volume of growth medium to inactivate trypsin after 2-3 minutes incubation
- 1.3.3 -centrifuge cells at 1000 RPM for 4min and resuspend in growth medium before adding to new flask
- 1.3.4 -cells should be passaged or fed at least twice per week

2. Assay Protocol

- 2.1 -Harvest cells from culture in growth medium and resuspend in assay medium
- 2.2 -dispense 4000 cells per well into 1536-well tissue culture treated white solid bottom plate using multidrop dispenser
- 2.3 -Incubate cells 5hr, then dispense 23nL of compound, positive control, or DMSO control using pintool
- 2.4 -positive and control compounds are located in the first four columns according to the plate map, and library compounds located in columns 5-48
- 2.5 -incubate plates for 19hr at 37 degrees C
- 2.6 -add 1uL of celltiter fluor to each well using bioraptr dispenser
- 2.7 -incubate 30min at 37 degrees C
- 2.8 -measure fluorescence using ViewLux
- 2.9 -Add 5uL of OneGlo using bioraptr dispenser
- 2.10 -incubate 30min at room temperature
- 2.11 -Read luminescence on ViewLux

3. Assay Performance

| AhR (Omeprazole) | Online Validation Agonist (Mean ± SD) | Online Validation Viability (Mean ± SD) |
|---------------------|---|--|
| EC50 | 33.2 ± 26.0 μM (n = 27) | NA |
| S/B | 4.83 ± 0.55 | 5.03 ± 0.16 |
| CV (%)* | 16.64 ± 4.29 (n = 18) | 7.76 ± 0.54 (n = 18) |
| Z' | 0.35 ± 0.09 | 0.69 ± 0.03 |

^{*} CV values shown represent average of DMSO plates and low concentration plates only.