

# Protocol of PXR-Luc HepG2 Cell-based Assay for High-throughput Screening

**DOCUMENT:** PXR-Luc HepG2\_TOX21\_SLP\_Version1.0  
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## ASSAY REFERENCES:

Assay Target	Cell Lines	Species	Tissue of Origin	Assay Readout	Assay Provider	Toxicity Pathway
Human PXR	HepG2	Human	Hepatocellular carcinoma	Luciferase reporter	Dr. Chen (St. Jude)	NR Signaling

## QUALITY CONTROL PRECAUTIONS:

1. -Maintain cells below 85-90% confluence.

## MATERIALS and INSTRUMENTS:

Supplies/Medium/Reagent	Manufacturer	Vender/Catalog Number
-EMEM	-ATCC	-ATCC/30-2003
-Phenol red-free DMEM medium	-Invitrogen	- Invitrogen/31053
-Fetal Bovine Serum	- Hyclone	- Hyclone/SH30071.03
-Charcoal/dextran treated FBS	- Invitrogen	- Invitrogen/12676
-Sodium pyruvate	-Invitrogen	-Invitrogen/11360
-L-Glutamine	- Invitrogen	- Invitrogen/25030
-Penicillin & Streptomycin	-Invitrogen	-Invitrogen/15140
- Geneticin (G418)	- Invitrogen	- Invitrogen/10131
-Recovery Cell culture Freezing Medium	-Invitrogen	-Invitrogen/12648
-0.05% Trypsin-EDTA	-Invitrogen	-Invitrogen/25300
-Rifampicin (Agonist control compound)	-Sigma	-Sigma/R7382
-DL-Sulforaphane (Antagonist control compound)	-Sigma	-Sigma/S4441
-Tetraoctyl ammonium bromide (Viability control compound)	-Sigma	-Sigma/294136
-1536-well white solid plates	-Greiner Bio-One	-Greiner Bio-One / 789173-

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-MULTIDROP COMBI	-Thermo Electron Corporation	-Thermo Electron Corporation
-BioRAPTR FRD	-Beckman Coulter	-Beckman Coulter
-ViewLux Plate Reader	-Perkin Elmer	-Perkin Elmer
-CellTiter-Fluor(TM) Assay System	-Promega	-Promega/G6082
-ONE-Glo(TM) Luciferase Assay System	-Promega	-Promega/E6130

## PROCEDURE:

### 1. Cell handling:

#### 1.1. Media Required:

Component	Growth Medium	Assay Medium	Thaw Medium	Freezing Medium
-EMEM	-90%		-90%	
-Phenol red-free DMEM medium		-95%		
-Fetal Bovine Serum	-10%		-10%	
-Charcoal/dextran treated FBS		-5%		
-Sodium pyruvate		-1mM		
-L-Glutamine		-2mM		
-Penicillin & Streptomycin	-100U/ml & 100ug/ml	-100U/ml & 100ug/ml	-100U/ml & 100ug/ml	
- Geneticin (G418)	-500ug/ml			
-Recovery Cell culture Freezing Medium				-100%

#### 1.2. Thawing method

1.2.1 -Thaw a vial of cells in 9ml of pre-warmed thaw medium and then centrifuge

1.2.2 -Re-suspend the pellet with the thaw medium and seed at 2 million cells per T-75 flask

#### 1.3. Propagation method

1.3.1 -Trypsinize cells from the culturing flask and centrifuge and then re-suspend cells in culture medium

1.3.2 -Passage cells at 2-3 million per T-225 flask

## 2. Assay Protocol

- 2.1 -Trypsinize cells from the culturing flask and centrifuge and then re-suspend cells in assay medium at a density of  $0.4 \times 10^6$  cells/mL
- 2.2 -Dispense 2000 cells/5uL/well into 1536-well tissue treated white/solid bottom plates using a 8 tip dispenser (Multidrop)
- 2.3 -Incubate the plates for 5hr at 37C and 5% CO<sub>2</sub>
- 2.4 -Transfer 23nL of compounds from the library collection (0.59nM to 92uM) and positive control through Pintool
- 2.5 -Incubate the plates for 24hr at 37C and 5% CO<sub>2</sub>
- 2.6 –After 23hrs of incubation at 37C, add 1ul of CellTiter-Fluor using single tip dispense (Bioraptr)
- 2.7 -Incubate the plates for 1hr at 37C and 5% CO<sub>2</sub>
- 2.8 –Read fluorescence (exposure time = 3sec) intensity using ViewLux plate reader
- 2.9 -Then add 4ul of ONE-Glo(TM) Luciferase reagent using a single tip dispense (Bioraptr)
- 2.10-Incubate the plates at room temperature for 30min
- 2.11- Read fluorescence (exposure time = 45sec) intensity using ViewLux plate reader

### 3. Assay Performance

Online Validation	PXR-Luc HepG2 Agonist mode	PXR-Luc HepG2 Viability-Agonist
CV*	$9.73 \pm 0.76$ (n = 21)	$4.33 \pm 0.45$ (n = 24)
S/B	$4.84 \pm 0.54$	$5.75 \pm 0.09$
Z	$0.66 \pm 0.07$	$0.85 \pm 0.02$

\*CV values shown represent average of all plates excluding high compound concentration plates.